

Dynamics of Glyphosate in the <u>Rhizosphere:</u> <u>A Possible Threat to Crop Plants?</u>



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Overview

- Introduction/Background
- Relevant knowledge on glyphosate
- Waiting times: An important issue?
- The Rhizosphere: Place for possible glyphosate toxicity
- Roots of target plants: Key players in stabilization and toxicity
- Conclusions / Prospects

Symposium on Mineral Nutrition and Disease Problems in Modern Agriculture: Threats to Sustainability?. Coplacana, Piracicaba, Brazil, 20.-21.Sept. 2007



Universität Hohenheim University Hohenheim (founded 1818)







Institut für Pflanzenernährung Institute of Plant Nutrition (founded 1923 for Prof. Margarethe v.Wrangell)







Fürftin Margarethe Andronitow Brangell D. 5. Professor an der Landwirtschaftlichen Hochschule Gründerin und Vorstand des Pflanzenernährungs Institutes im Treibhaus ihres Institute, 1925



ICH LEBTE MIT DEN PFLANZEN ICH LEGTE DAS OHR AN DEN BODEN UND ES SCHIEN MIR ALS SEIEN DIE PFLANZEN FROH ET WAS ÜBER DIE GEHEIMNISSE DER RHIZOSPHÄRE ERZAHLEN ZU KONNEN

(1876 - 1932)

Main research interest: 4 Role of rhizosphere processes in P acquisition of P-efficient plant species



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Introduction/Background



Glyphosate:

- Worldwide the most widely used herbicide (Trade name "Roundup).
- Non-selective, inhibits synthesis of aromatic amino acids via the shikimate pathway.
- Efficient and cheap low production costs
- ➢ General claimed (e.g. by Monsanto) :
- rapid microbial degradation and / or binding to the soil (= detoxification)
- no residual effects in soils
- no negative environmental effects

However, recent observations suggest significant side effects on non-target organisms!!

Observed interactions between glyphosate and crop plants

• Partial desiccation of cover crops after wheat by accidental double application of glyphosate (4L/ha glyphosate) before sowing of cover crops



(Farm near Tübingen, Germany 2006)

• <u>Enhanced drought stress after glyphosate applications</u> (see:glyphosate case between cotton growers in Texas and Monsanto)

> due to strongly inhibited root growth or to impeded nutrient acquisition (Mn, Zn, Fe, K) and thus due to more heat stress problems.

• Drought stress partially linked with enhanced root diseases



Drought spells in sugar cane due to take-all (Sao Paulo State, 2004)

Greening effect of `Weather Max', a new formulation of glyphosate by Monsanto, as observed by Myriam Fernandez in the field: a positive effect of glyphosate?

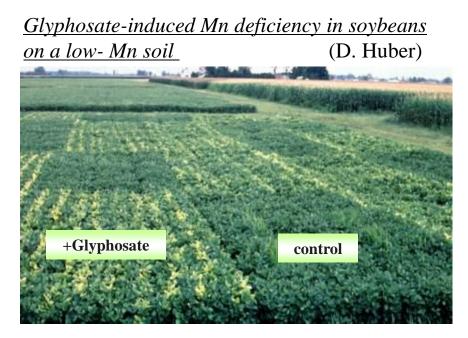
- By `Weather Max' Myriam observed a better performance of the following crop (taller plants, more green, delayed senescence, higher plant N and lower S concentration)
- (Possible explanation: Release of soluble N from microbial biomass after short-term killing of soil microorganisms by glyphosate; immobilization of S by the flush of easily plant available N). <u>No long-term positive effect for sustainability!?</u>

Observed interactions between glyphosate and micronutrients

In the USA with a high percentage of RR (Roundup-resistant)-crops, there are increasing reports on:

- micronutrient deficiencies induced by glyphosate
- increase in demand for micronutrient foliar fertilizers

(Jurin, 2004; Brown, 2005)



Interaction of seed applied Fe and glyphosate application on Fe deficiency chlorosis in soybeans; Minnesota, USA (Jolley et al., Soil Sci Plant Nutr. 50, 793-981, 2004)

V	isual chloro	sis scrore	grain	yield	Treatment
(1=green to $5=$ severe $)$		(t/ha)			
– Fe	+ Fe*	– Fe	- + Fe*	:	
Control (no herbicide)	3.1	2.8	1.01	1.70	
Glyphosate	3.7	3.3	0.27	0.61	

* 50g Fe/ha as FeEDDHA applied to seeds

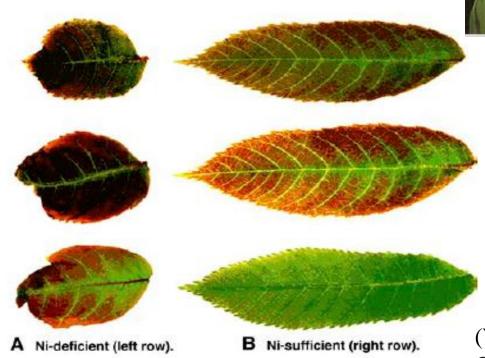
Ni deficiency in pecan trees:

glyphosate-induced similar to Mn- and Fe-deficiency as assumed by Yamada? -via strongly inhibited root growth by glyphosate,

- via inhibited micronutrient acquisition and thus susceptibility to heat stress,

(- besides high Zn-induced Ni deficiency).

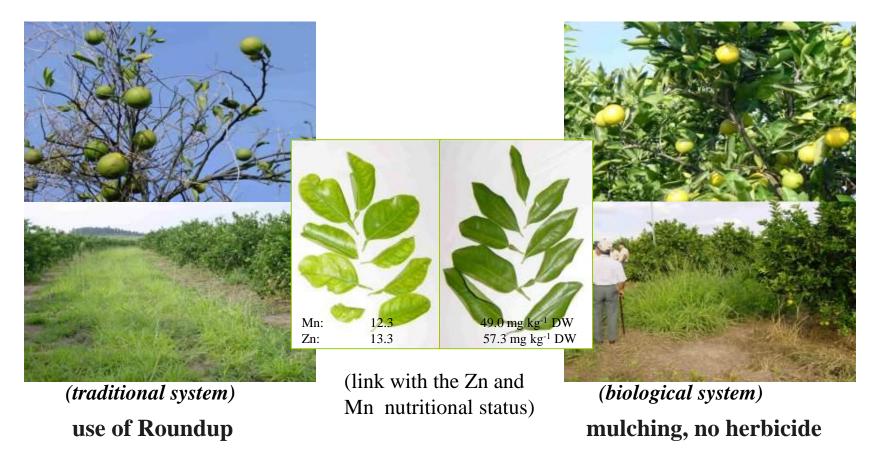




(Wood et al. 2003; Chen Bai et al. 2006)

Observed interactions between glyphosate and diseases

The <u>dieback syndrome (C.V.C.)</u> is particularly expressed in traditional production systems with a high application rate of the herbicide Roundup (Glyphosate), but less in biological production systems with *Brachiaria* mulch for weed control.





High incidence level of *Fusarium* Head Blight (FHB) in wheat in Saskatchewan, Canada



"Risk Production Factors" associated with FHB:

Environment (rainfall, temperature)

Crop Production Factors-

** Roundup applied 18-36 months prior to wheat planting had the <u>most</u> <u>consistent</u> relationship to FHB development across all years studied.

Fernandez et al., 2005; Crop Sci. <u>45</u>, 1908-1916

<u>A wide range of observations believed due to glyphosate</u> <u>applications:</u> How can they all induced by glyphosate or explained?

- <u>What do we know on glyphosate</u> for understanding these various before mentioned observations in fields?

In discussions with various representatives of Monsanto (e.g. Brazil, Europe, St Louis USA)

<u>no</u> links between these mentioned observations and glyphosate use!

Safety, always and everywhere!

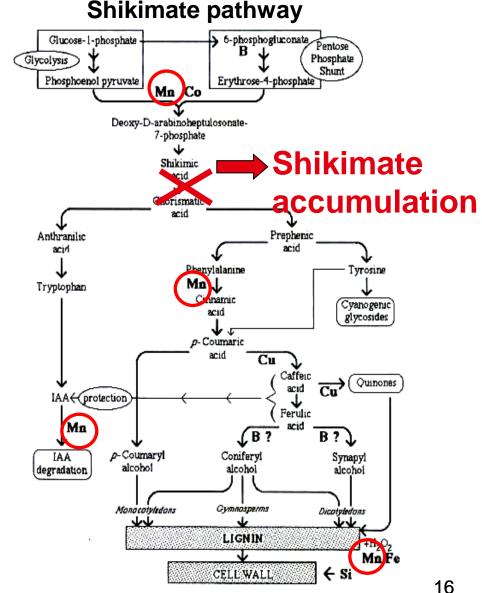
- However: <u>What have we to know on glyphosate</u> for a better understanding <u>and possible counteraction</u> against these observed negative effects by management?
- Need for a more integrative or holistic view!

Relevant knowledge on glyphosate

Glyphosate is a non-selective, systemic, phloem-mobile inhibitor of the enzyme EPSPs, disrupting the shikimate pathway for biosynthesis of essential aromatic amino acids such as tryptophan, phenylalanine and tyrosine.

In plants, glyphosate is quite stable, with little detectable degradation occurring over long periods and tends to accumulate in the meristematic regions.

Source: Gruys & Sikorki, (1999).



□ <u>Relevant knowledge on glyphosate</u>

Strong fixation to soil = immobilization = detoxification

(possible re-mobilization as a phosphoric compound?)

Inhibition of the shikimate pathway (see presentation before!)

Preferential transport within target plants to apical tissue (e.g. root tips)

<u>Release into the rhizosphere</u> (*scheme of overlapping rhizosphere of a target and non-target plant root*!)

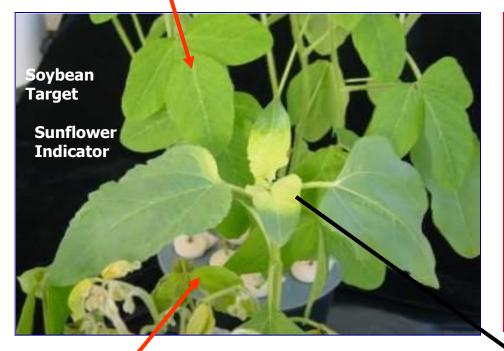
<u>and what is then</u>? What is the mechanism of this release into the rhizosphere and how fast is this release depending on which factors?

(important questions which are not seriously adressed by Monsanto or even by S. O. Duke as a well-known herbologist from USDA, $_{17}$ USA)

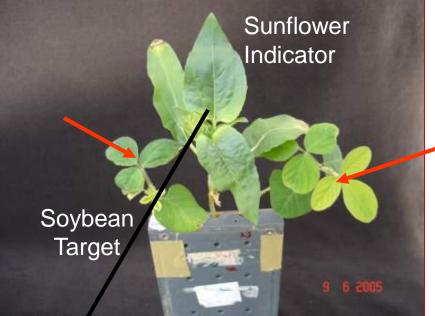
<u>Glyphosate applied to target plants (weed) can be released into</u> <u>the rhizosphere</u>

Induction of Fe deficiency chlorosis in non-target plants (sunflower) induced by glyphosate transfer from foliar treated target plants (soybean)

Nutrient solution experiment



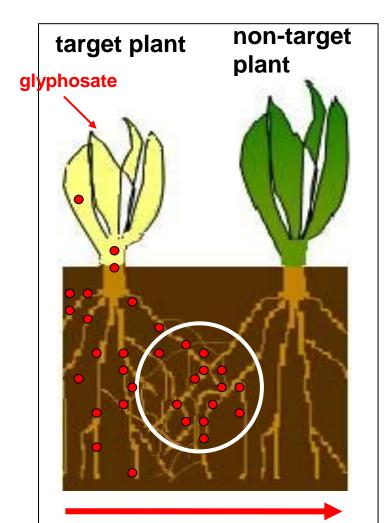
Rhizobox experiment



Glyphosate application to target plants

Fe deficiency symptoms in nontarget plants ... and accumulation &f shikimate!

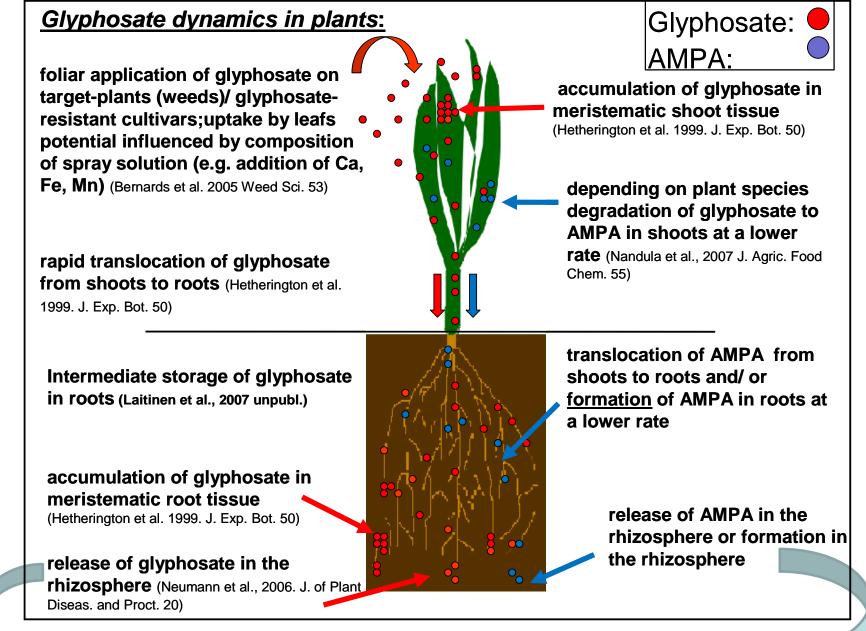
□ <u>Relevant knowledge on glyphosate</u>



Glyphosate-transfer via shared rhizosphere

What have we to know?

After accumulation of glyphosate in the roots of target plants (e. g. weed) release into the rhizosphere with possible consequences for a nontarget crop plant!



Open questions: What is the mechanism of this release into the rhizosphere and how fast is this release depending on which factors?

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□ <u>Relevant knowledge on glyphosate</u>

• How long this toxic glyphosate or AMPA can be stored in roots of target plants..... depending on which soil and management factors?

Important questions for the issue of waiting times after glyphosate use by farmers!

Waiting times: An important issue?

Regarding Monsanto's representatives (2006) there is <u>no need for</u> <u>waiting times</u> to be considered! No need for such an indication on package label for directions for use by farmers!

Even advertisement for an use of glyphosate till one week after sowing in Germany or Brazil!

Aus Erfahrung besser!



<u>Is this general statement of</u> <u>Monsanto responsible to farmers</u> <u>and in agreement with increasing</u> <u>observations by farmers and</u> <u>research result during the last</u> <u>years?</u>

Time of	Cover crop			
	Black oat	Ryegrass	Fallow	
21 dbp	(100)	(100)	(100)	
14 dbp	-2.1	-7.3	-3.7	
7 dbp	-6.8	-18.5	-12.3	
0	-11.2	-23.4	-17.2	
7 dap	-17.4	-25.9	-21.2	

Effects of timing of cover crop desiccation on RR soybean yield

dbp = days before planting; dap = days after planting

(Aroldo Marochi, 2006)

Clearly, best time for glyphosate application <u>2-3 weeks before sowing</u> the following crop (even for RR soybeans) in Brazil on low buffered soils!

Results by POTAFOS, Brazil showing the need of waiting times



dap = day after planting, dbp = days before planting

"best plant development when sowing soybean 14-21 days after desiccation by glyphosate"

<u>Relevance of waiting times after weed glyphosate desiccation</u> (model green house experiment) :

Luvisol



Sunflower plants grown on a Luvisol (subsoil) sown 0, 7, 14, 21DAA (after glyphosate application) to weed or mechanical weeding (-Gly).

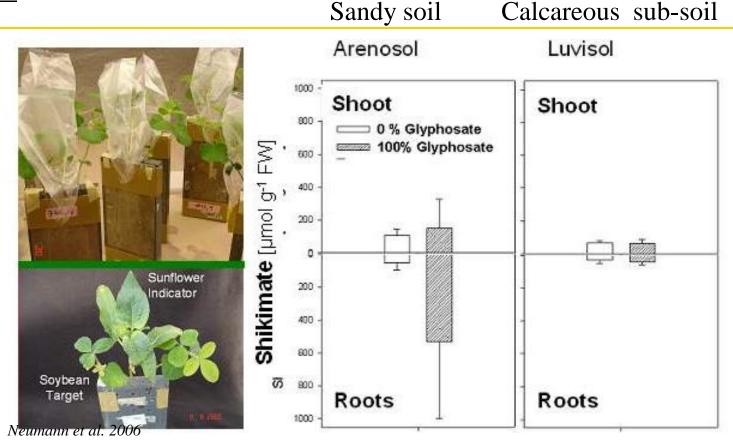


Sunflower plants grown on an Arenosol sown 0, 7, 14, 21DAA (after glyphosate application) to weed or mechanical weeding (-Gly).

Sever plant growth inhibition if waiting time is less than 21 days and a stronger observed toxicity if buffering capacity of the soil is low.

This indicates relevance of waiting time in glyphosate use and the consideration of the soil type!
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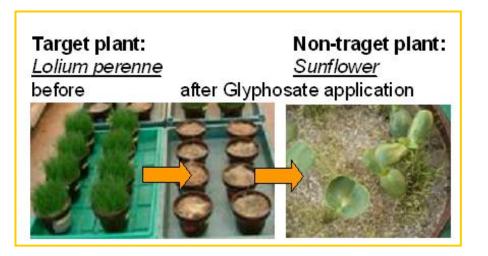
Soil type dependent **Short-term rhizosphere transfer** of glyphosate from glyphosatetreated RR soybean (recommended dosage) to simultaneously cultivated, untreated <u>sunflower.</u>



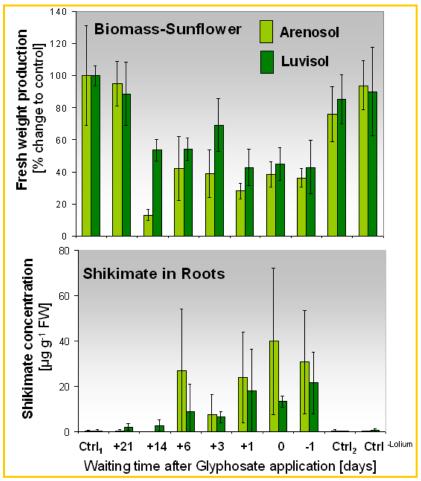
Shikimate accumulation (indicator for glyphosate toxicity) in sunflower **<u>7 days after glyphosate application</u>** to soybean

Glyphosate-induced shikimate accumulation in non-target sunflower plants on the Arenosol, <u>but not</u> on the calcareous soil (rapid immobilisation of glyphosate on the calcareous soil as Ca-salts ???)

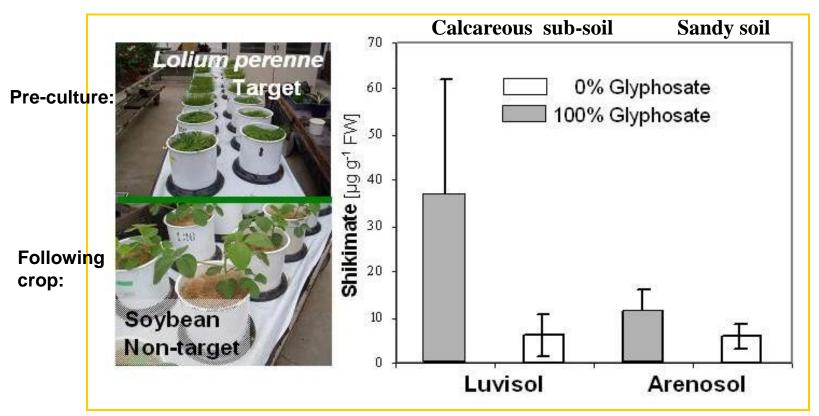
Root to Root transfer of glyphosate from target (*Lolium perenne*) to non-target plants (sunflower) depending on waiting time after glyphosate application



By waiting time of less than 14 days inhibited shoot growth and shikimate accumulation in roots! Plant growth and intracellular shikimate accumulation as physiological indicator for glyphosate toxicity.



Long-term rhizosphere transfer from glyphosate-treated Lolium perenne to simultaneously cultivated untreated soybean.



Shikimate accumulation (indicator for glyphosate toxicity) in soybean **<u>8 weeks after glyphosate application</u>** to *Lolium perenne*

Glyphosate-induced shikimate accumulation in non-target plants on the calcareous soil (re-mobilisation of fixed glyphosate?) but not on the Arenosol with low glyphosate immobilisation (complete microbial degradation within 8 weeks?)

ראונדאפ קוטלי ROUNDUP* עשבים הדברת עשבים לקראת גידולי שדה ובשטחי כרב המינון העשבים מועד הטיפול הגידול ('T/7"DO) המודברים לפני הזריעה או לאחריה. **DOINT 100** א, נבטי חד-שנתיים עד גובה לקראת גידולי קיץ: וכן לקראת שתילה. DX 1000 0.5% סו סימ, לרבות נבטי דורת כותנה, מקשה, אגוד אין לרסס לאחר השקיית ארם-צובא שהציצו מזרעים. אדמה, תירס ורקות. ההנבטה, למעט בקרקעות חסטותם 150 .010 40-10 D*D0-10.2 חוליות המכילות פחות שפ.ס משטח XD יבלית ודורת ארם-צובא מקני מ־201 חרסית. שורש ייצרבו וידוכאו זמנית. עשבים הב-שנתים: יבלית, דורת 500 (במוקדים 5%) במצע מזכן לפני הזריעה לקראת זריעת גידולי ולאחריה (לא לשתילה). ארם-צובא, חבלבל, גומא אין לרסס לאחר השקיית הפקעים-בעלי עלווה מפותחת. כותנה, מקשה, אגוזי ההנבטה. למעט בקרקעות אדמה, תירס וירקות. ססג (במוקדים 3%) חוליות המכילות פחות דורת ארם-צובא בלבד. מ'%01 חרסית. על שלה ועשבים מלבלבים 200 TV 150 שלף מתחדש ועשבים לקראת כותנה לאחר השקית יסוד וזריעת בהתאם לגובה חר-שנתים קיימים, לרבות בדו-גידול בזריעה הכותנה, לא יאוחר מיומיים העשבים, בתוספת למישרין (לאחר חיסה, דורת ארם-צובא שהציץ לפני הצצתה. DX 10000 0.5% מזרעים. שעורה, שחת). על כרב אז לאחר פנוי פני 500 TV 300 טיפול קיצי לקראת עשביה רב-שנתית הקרקע, משאריות הגידול 150-100 JODIJJ הכוללת יבלית, גומא דגני חורף בלבד. הקורם, כל עוד העשבים טומהוק (מינון הפקעים, דורת ארם-צובא, ראונדאפ הגבוה בצימוח פעיל. ינבוט, חבלבל, הגא, להדברת יבלית וגומא הפקעים)* 500 יבלית בעלת עלווה מפותחת (במוקדים 15% וכן עשביה אחרת הקיימת LIGH RPOID. * בריסוס בתערובת ראונדאפ עם אלבר סופר או טומהוק, עלול רחף התרסיס לגרום לפגיעה חמורה בגידולים רגישים לתכשירים אלה, לכן השימוש בשילוב זה כפוף להנחיות ולאמצעי הזהירות בעת הריסוס המפורטים בתויות, לרבות ההוראות לניקוי המרסס לאחר הטיפול. נפח התרסיס: בריסוס קרקע 15-10 ל׳/ד׳. בריסוס אוירי 5 ל׳/ד׳ (ראה תוית). טיפול בראונדאפ להקמלת הגידול נפח המינון מועד הטיפול הגידול התרסיס ('oa"ה/ר') L'010 20-10 -02 4'/T עד דימים לפני הקציר 300-100 חיטה, שעורה, בקיה, ש"ש, תירס, ריסוס אוויר: 5-7 לי/ד פנסילריה-לשחת או לתהמיץ הערה: אין לטפל בגידול המיועד לזרעים.

In Israel: Glyphpsate use on dry and sandy soils forbidden as mentioned on the package label for farmers use.



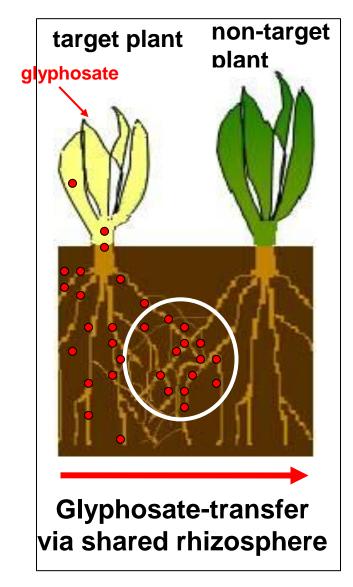
The results by Myriam Fernandez on negative effects of glyphosate on FHB incidence in Canada even 18-36 months after glyphosate application might indicate <u>even longer waiting times</u> in distinct situations with a long lasting glyphosate effect! <u>In conclusion</u>, waiting times after weed control with glyphosate might be

- 0 3 weeks for wet, light soils with a fast turn-over of weed roots (e.g. in Brazil),
- 4 8 weeks for wet, heavy calcareous soils with a slower turn-over of weed roots,

but might be up to

- 1 year for dry sandy soils as wide-spread in Israel,
- 1.5 3.0 years for cold soils with an impeded turn-over of weed roots as in some regions of Canada.

The Rhizosphere: An important place for possible glyphosate toxicity



Obviously, various <u>processes</u> of glyphosate dynamics take part in the immediate vicinity of roots, the so-called <u>rhizosphere.</u>

What are these various processes of importance for glyphosate toxicity?

□The Rhizosphere: An important place for possible glyphosate toxicity

glyphosate dynamics in the rhizosphere:

chemical processes:

chemical degradation: (e.g. by metal oxides)

Stabilization of glyphosate in the rhizosphere:

low degradation: •low activity of MOs

low fixation

- high P
- · low clay
- · low divalent cations

Fixation of

<u>glyphosate in soils:</u> Ca, Fe, Al, clay, OM, Iow P

Morillo et al.2000, Gimsing et al. 2004 Sörensen et al. 2006 translocation of glyphosate from shoot to root and subsequent release into the rhizosphere:

potential consequences:

formation of phyto-toxic metabolits (e.g. AMPA)

<u>Transfer of glyphosate</u> <u>to non-target plants,</u> glyphosate-toxicity, damage, yield reduction

Reduced availability of microntrients (Fe, Zn, Mn)

<u>Over time:</u> Potential long-term storage and remobilization

Leaching of Glyphosate:(Kjæret al. 2005)

□The Rhizosphere: An important place for possible glyphosate toxicity

beneficial MOs

pathogens

glyphosate dynamics in the rhizosphere (cont.)

biological processes:

biologial degradation: Glyphosate as C, N or P source for microorganisms

Glyphosate-toxicity:

Inhibition of bacterial shikimate pathway

Toxicity on mycorrhiza

translocation of glyphosate from shoot to root and subsequent release into the rhizosphere:

Potential consequences formation of phyto-toxic <u>metabolits (e.g. AMPA)</u> promotion of pathogenic <u>microorganisms (e.g.</u> Fusarium, Pythium-species) Higher risk of diseases promotion of Mnoxidizing microorganisms Mn-deficiency, effect on defence mechanisms of <u>plants</u>

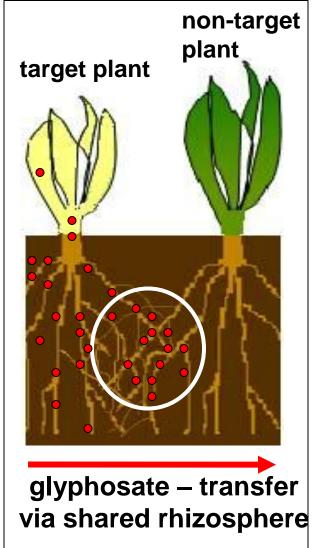
<u>Less Mn (Mn-reducers)</u> <u>Less N (impaired BNF)</u> <u>Less P (Mycorrhiza)</u>

□The Rhizosphere: An important place for possible glyphosate toxicity

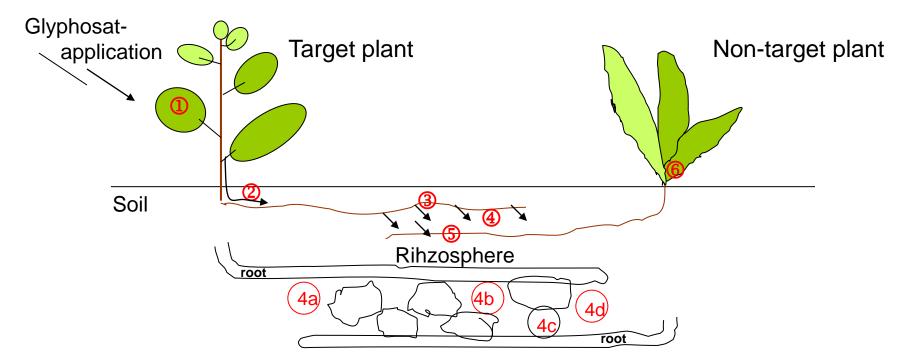
These processes and their interdependencies may change with:

- soil chemical properties (pH, redox)
- microbial population
- application frequency
- application time
- plant species
- over time

The role of the rhizosphere as place for glyphosate toxicity may drastically increase in case of a shared rhizosphere between glyphosate treated and non-treated plants



Dynamics of Glyphosate/AMPA in the Rhizosphere (Model)



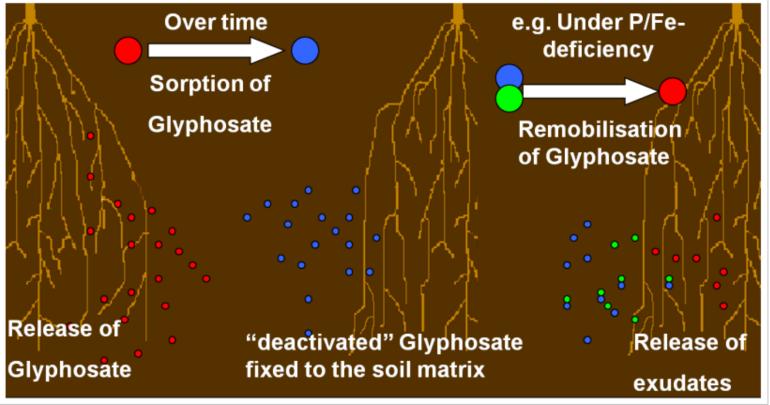
- ① foliar uptake of glyphosate
- ^② transfer of glyphosate into apical root zones
- release of glyphosate and possible metabolites (AMPA) into the rhizosphere of target plants or degradation of root residues
- ④ glyphosate dynamics in the rhizosphere
- ⑤ uptake of glyphosate by non-target plants
- © translocation of glyphosate/AMPA into the shoot of non-target plants and induction of disorders

- ④ glyphosate/ AMPA dynamics in the rhizosphere
- a) extent of interactions between root system of target and non-target plants (intermingled roots)
- b) glyphosate immobilization in the rhizosphere
- c) glyphosate remobilization by root-induced changes in the rhizosphere of non-target plants
- d) interaction of glyphosate with Mnreducing/oxidizing rhizosphere microorganisms
- e) effect of glyphosate on mycorrhizae and ³⁹ microbial diversity

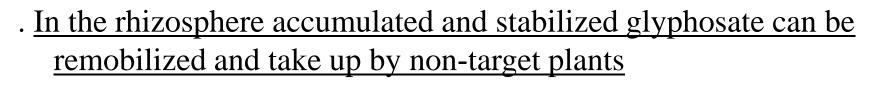
Remobilization of glyphosate from soils: a possible reason for prolonged glyphosate-toxicity in soils?

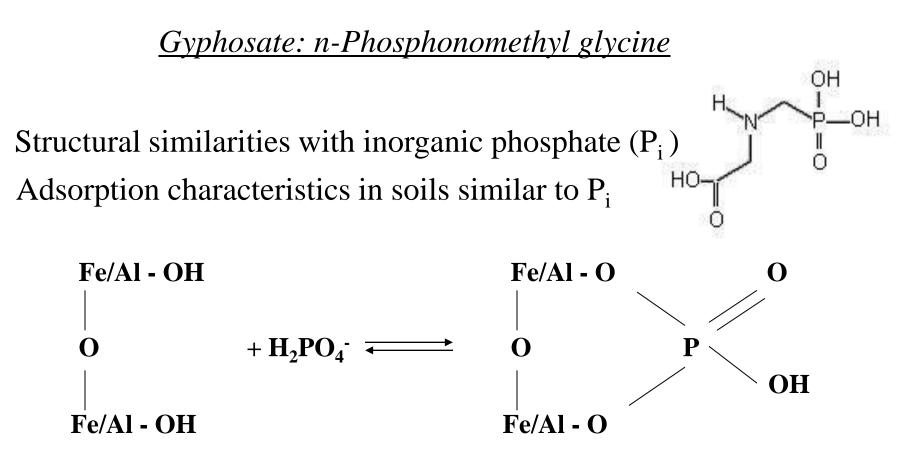
In soils, glyphosate behaves similar to P by strong adsorption to Fe, Al, Ca, organic matter and clay minerals (Morillo et al., 2000, Gimsing et al., 2004, Sörensen et al., 2006)

BUT: a remobilization e.g. by carboxylates released under nutrient deficiency has to be considered!



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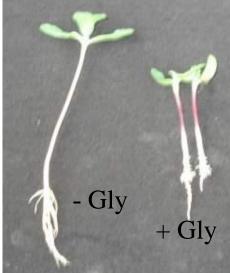




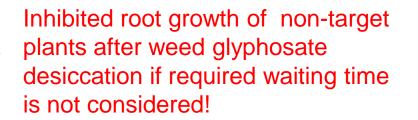
From these consideration it can be concluded that P-efficient plant species will mobilize glyphosate more efficient under low P status and that measures for a better P fertilizer use (e.g. pH lowering, silicate or water-soluble humic 41 substances) will also enhance a remobilization of glyphosate !

Roots of non-target plants as prime victims of

glyphosate residual toxicity:



Sunflower seedlings grown on an acidic Arenosol 14 days after glyphosate weed (*Lolium perenne*) desiccation.

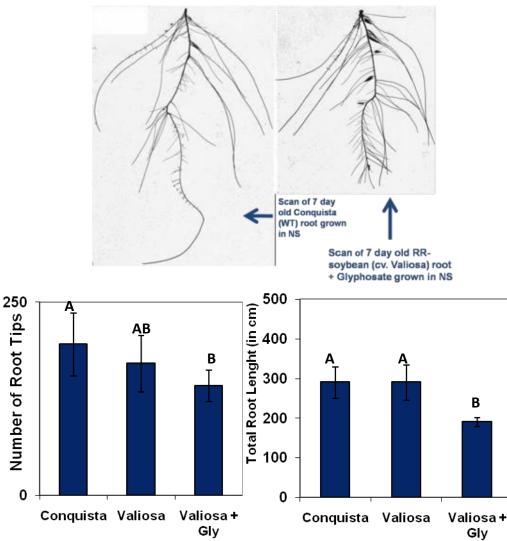




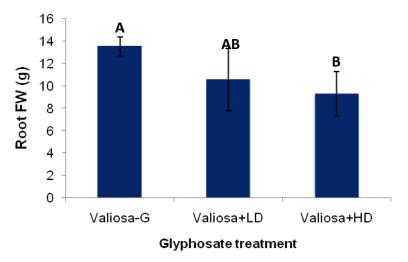


Sunflower roots grown on an acidic Arenosol (top) and calcareous Luvisol sub soil (bottom) at 0 days waiting time after glyphosate desiccation of pre-cultivated weed 43

<u>Glyphosate effect on root morphology and -growth of RR soybean</u> plants (cv. Valiosa) grown in soil and nutrient solution cultures.



Reduced root elongation 4 days after 28.4mM Glyphosate (recommended rate) application to RR soybean (cv.Valiosa) grown in hydroponics (formation of shorter and reduced number of roots).

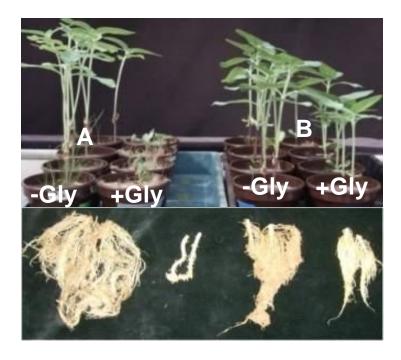


Inhibition of root biomass of RR soybean (cv. Valiosa) grown on calcareous soil due to glyphosate application at lower (LD i.e.2L/ha) and higher (HD i.e. 4L/ha) range of recommended dosage proposed by the producer company.

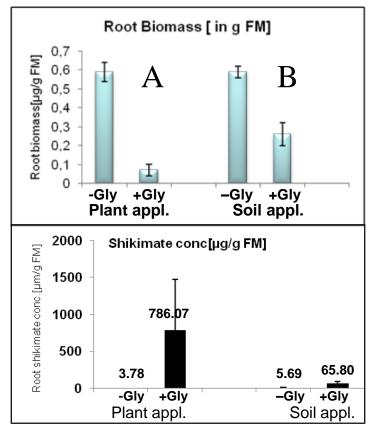
Results on root growth and morphology of nontarget and RR-plants highlight risk of increased drought stress by 44 glyphosate use.

Roots of target plants: Key players in affecting stabilization and toxicity of glyphosate

(Green-house model experiment)

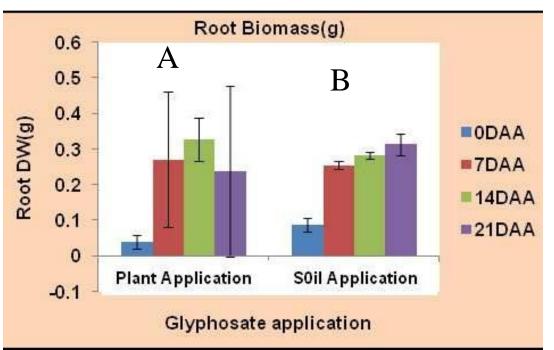


Inhibited sunflower seedling growth (both shoot and root) sown zero days after glyphosate desiccation of precultivated Lolium perenne as weed(A) and direct soil application(B). Stronger effect in weed (A) than soil application (B)!



Root biomass and intracellular shikimate accumulation of sunflower seedlings grown 0 days after Lolium perenne weed glyphosate desiccation (plant appl.) and direct soil incorporation (soil appl.). Stronger residual toxic effect in plant (A) than soil application (B). 46

Differential pattern of glyphosate residual toxicity between target plant application and direct soil application



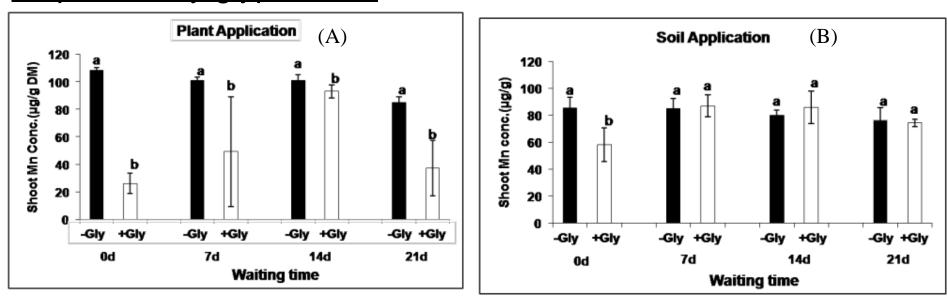
Extended glyphosate residual toxicity after plant application compared to soil incorporation.

Root biomass of sunflower plants grown on acidic Arenosol after glyphosate Lolium perenne weed desiccation or direct soil application.

Note: The big standard errors in plant application (A) seem to represent hot spot glyphosate pool formation in the rhizosphere rather than due to random sampling variability, as there were similar high differences in plant growth within the same pot.

Similar hot spot effects of gyphosate were observed in Ni deficiency of pecan trees by Wood et al. and Bai et al. (see before).

Soil type and application mode dependent inhibition of Mn acquisition by glyphosate: Arenosol



Mn concentration of sunflower plants grown on <u>acidic Arenosol I with low buffering capacity</u> at different waiting times after *Lolium perenne* weed glyphosate desiccation (plant application) and direct soil incorporation (soil application).

In soils with a low buffering capacity, glyphosate residual toxicity can be extended up to 21 days waiting time!!



Soil type dependent role of roots in stabilization process of glyphosate in a soil!!.

□Roots of target plants: Key players in affecting stabilization and toxicity of glyphosate

Clear indications for roots as key players in stabilization of glyphosate in pot experiments with sunflower, BUT: These findings of the model pot experiment <u>need further</u> <u>confirmation by</u>

- further distinct pot experiments and
- field experiments with different crops (on-going!)

Further, the research of the Italian group (Senesi et al. 199x) on the stabilization of glyphosate on organic matter in the rhizosphere (root exudates?) has to get re-examined including the turn-over of weed roots, high in accumulated glyphosate.

Conclusions / Prospects

- Farmers of <u>non-till practice</u> in Brazil are <u>in favor of glyphosate</u>.
- However, they recognize <u>increasing problems with micronutrient deficiencies</u>, <u>drought and disease problems</u>.
- With innovative rotations (including black oat), higher micronutrient fertilization and more pesticide application they try to counteract at least partially these problems.
- For a better understanding of the non-foreseen negative side-effects of glyphosate by Monsanto <u>the rhizosphere as the immediate vicinity of roots has</u> to be taken into consideration.
- Obviously, <u>in the earlier studies</u> with a rapid detoxification or immobilization of glyphosate in soils, <u>the rhizosphere of target (weed) plants was not properly considered.</u>
- Glyphosate and its high toxic metabolite AMPA (amino-ethylphosphonic acid), released into the rhizosphere of target plants are <u>long enough stable to be taken</u> <u>up by following crop plants</u> (non-target plants) <u>with detrimental effects if</u> <u>waiting times are not considered.</u>

Conclusions / Prospects (continuation)

- Roots of target (weed) plants are the key players affecting stabilization and toxicity of glyphosate depending on the conditions of degradation of the glyphosate containing root residues (soil type and weather dependent).
- A possible re-mobilization of soil-adsorbed glyphosate in the rhizosphere of nontarget plants after repeated application of the herbicide over the years, particularly under non-till practice, is not seriously considered up till now.
- <u>A new risk assessment</u> for glyphosate including the rhizosphere processes with stabilized glyphosate in root residues is urgently claimed, in particular if the expected increasing use of Roundup-resistant (RR) cultivars world-wide is considered.
- To avoid negative effects of glyphosate on plant growth and micronutrient acquisition and thus on disease resistance of the following crop, the turnover of glyphosate in the rizosphere via an adequate waiting-time for different soil types and weather conditions have to get elaborated.
- For all the above mentioned requests a stop of the highly polarized or black and white discussion of the glyphosate issue is urgently needed! 52



A <u>modeling approach</u> might help to <u>predict the</u> <u>needed waiting time</u> to avoid negative sideeffects of glyphosate depending on conditions

depending on conditions for degradation and thus release of stabilized glyphosate in roots of target (weed) plants as key players in glyphosate toxicity in the rhizosphere.

Muito obrigado! Thank you for your kind attention!

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